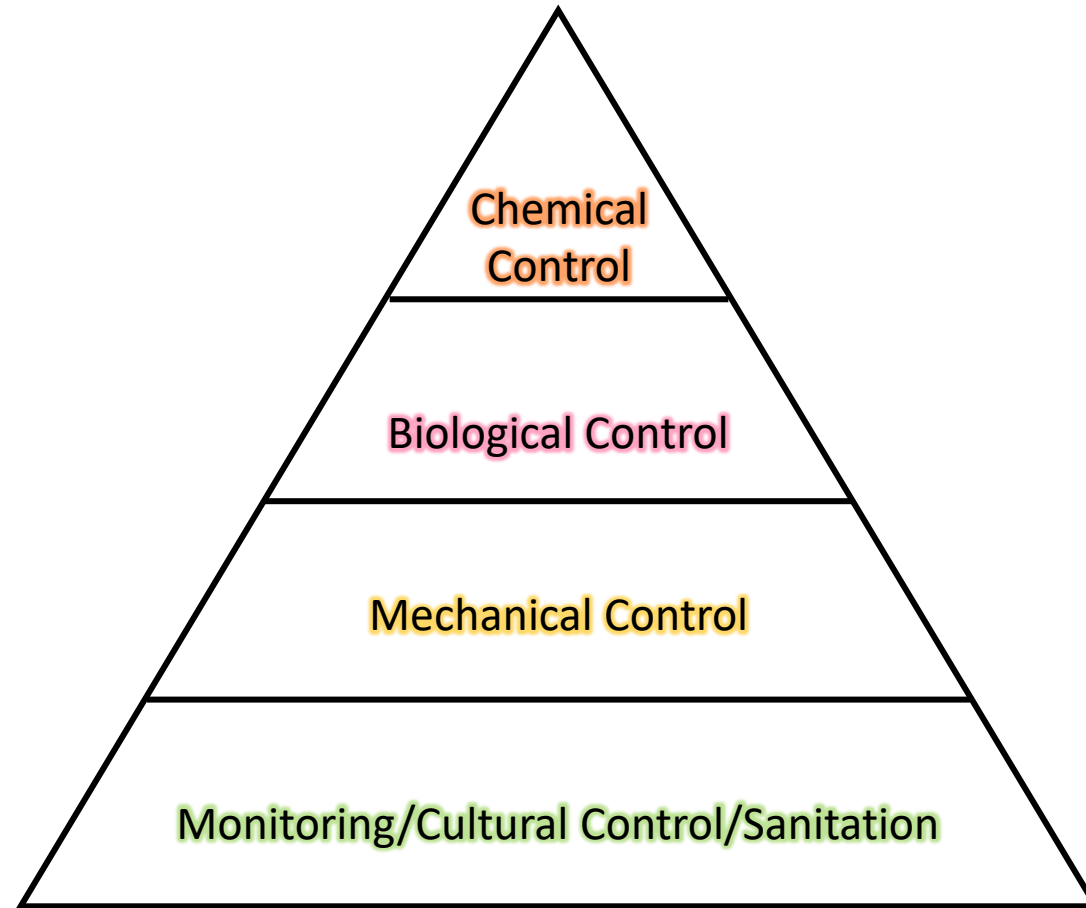


# The Hibiscus Bud Weevil: Biological Control & Sanitation Practices

Alexandra M. Revynthi

German Vargas, Yisell Velazquez Hernandez, A. Daniel Greene, Isamar Reyes-Arauz, Maria A. Canon

# Integrated Pest Management



# Chemical Control Against HBW- Contact Insecticides

- No use of neonicotinoids (4A)
- We evaluated 17 insecticides and 4 oils
- The most efficacious were:

Product	Active Ingredient	Group	Rate
Dimilin 25 W	diflubenzuron	15	16 oz/100 gal
Xxpire	sulfoxaflor+spinetoram	4C + 5	0.08 oz/3 gal
Pyganic EC	pyrethrins	3A	15.61 fl oz/100 gal
Kontos	spirotetramat	23	0.1 fl oz/3 gal

# Chemical Control Against HBW- Systemic Insecticides

- No use of neonicotinoids (4A)
- We evaluated 4 insecticides in a prophylactic and curative manner

Product	Active Ingredient	Group	Rate
Altus	flupyradifurone	4D	28 fl oz/750 gal
Mainspring	cyantraniliprole	28	12.2 fl oz/100 gal
Acelepryn	chlorantraniliprole	28	16 fl oz/100 gal
Kontos	spirotetramat	23	0.1 fl oz/3 gal

# Biological Control of the HBW

- Entomopathogenic:
  - Nematodes (EPNs)
  - Fungi
  - Bacteria
  
- Parasitoids

# Biological Control of HBW - EPNs

Tested entomopathogenic nematodes (EPNs) in lab assays against L1, L2 and L3

- *Heterorhabditis bacteriophora*
- *Steinernema carpocapsae*
- *S. kraussei*
- *S. feltiae*
- *S. riobrave*

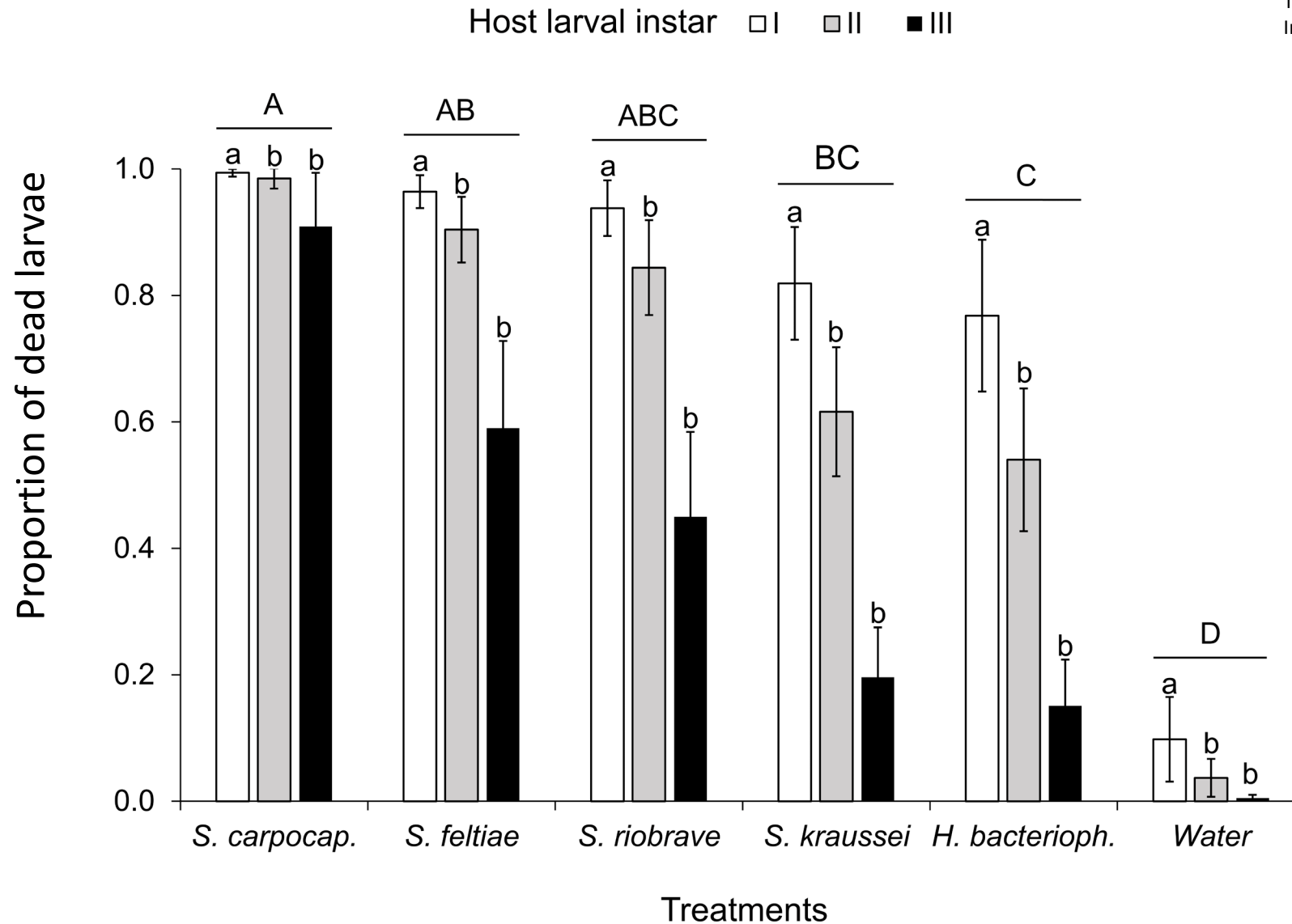
Rate:

500 IJ/bud approx. on top of naturally infested buds



# Laboratory Evaluations - EPNs

Treatment: ( $\chi^2 = 44.20$ ;  $df = 5$ ;  $P < 0.001$ )  
 Instar: ( $\chi^2 = 22.50$ ;  $df = 2$ ;  $P < 0.001$ )



# Greenhouse Evaluations

**Infestation:** 2 pairs of HBW

- *Steinernema carpocapsae*
- *S. feltiae*
- Water control
- w/ and w/o Barricade® (0.3%)

**Dosage:**

500,000 IJ/m<sup>2</sup> approx. sprayed at the foliage until runoff

N =6 plants per treatment and nematode combination

**Observations:** 5 actively growing buds and 5 fallen buds were dissected 1wk after the application

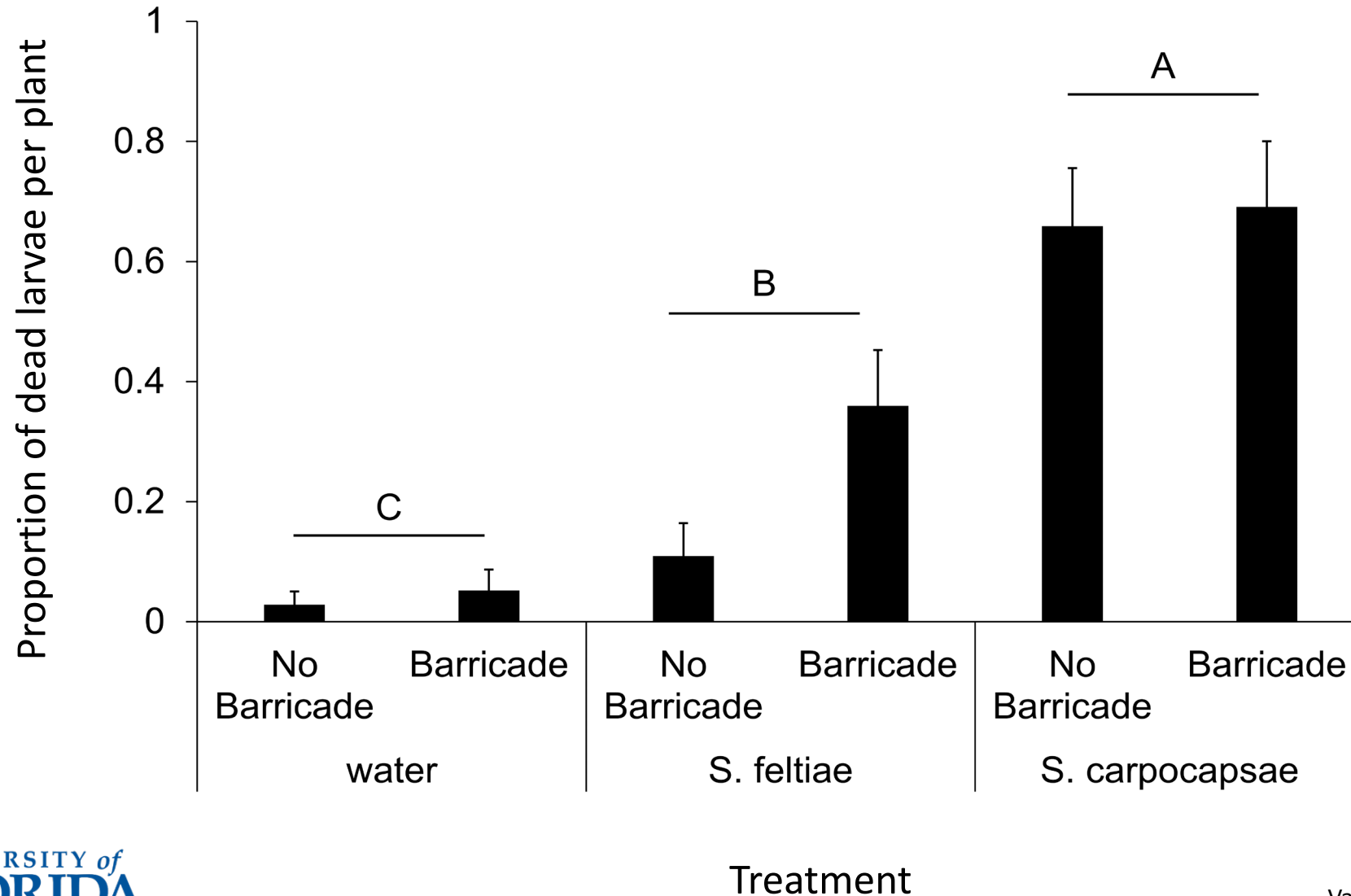


Photo: G. Vargas



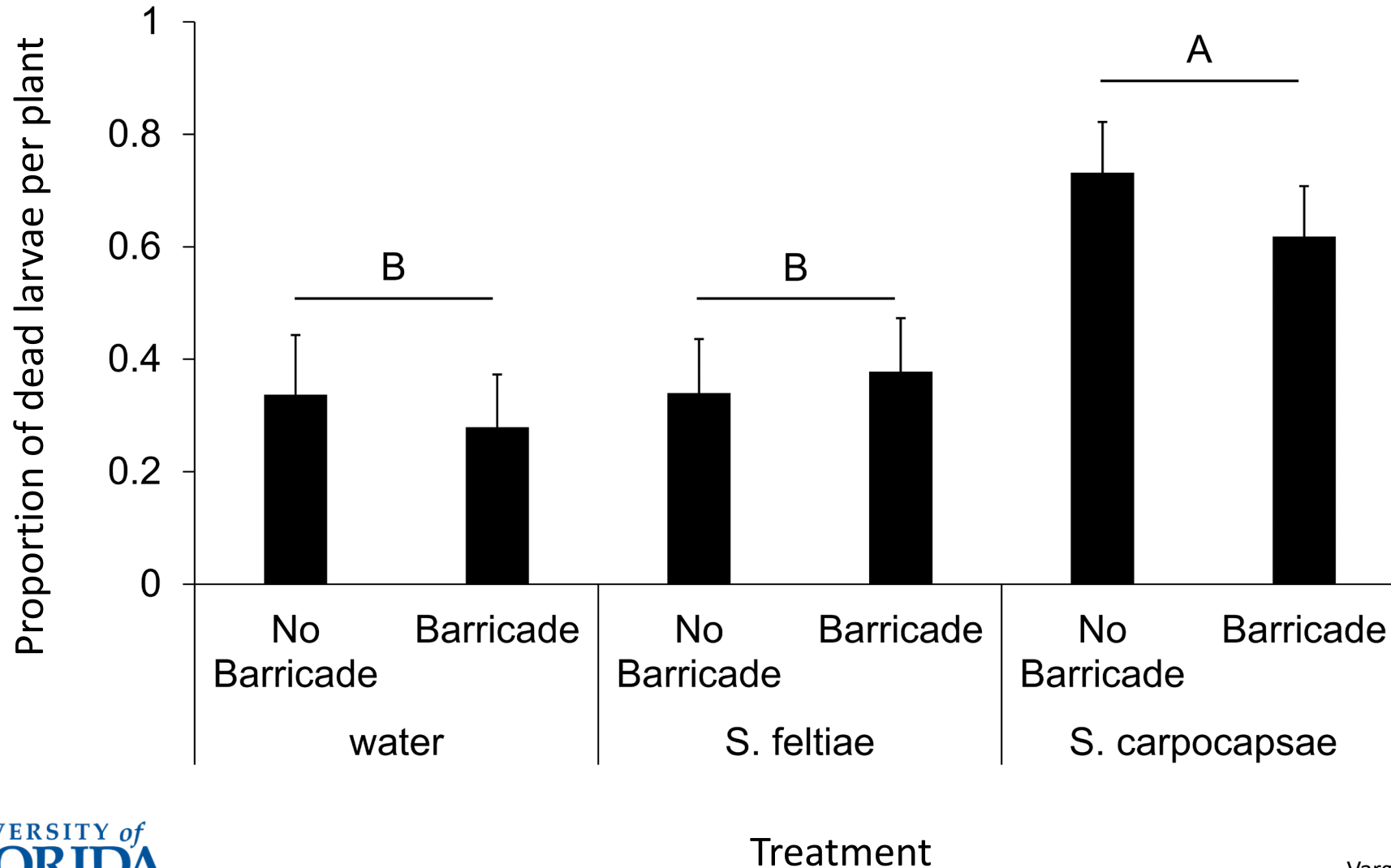
# Greenhouse Evaluations- Actively Growing Buds

EPN: ( $\chi^2 = 44.53$ ;  $df = 2$ ;  $P < 0.001$ )  
 Barricade: ( $\chi^2 = 3.7$ ;  $df = 1$ ;  $P = 0.054$ )



# Greenhouse Evaluations- Fallen Buds

EPN: ( $\chi^2 = 15.39$ ;  $df = 2$ ;  $P < 0.001$ )  
Barricade: ( $\chi^2 = 0.393$ ;  $df = 1$ ;  $P = 0.564$ )



# *Steinernema carpocapsae*



# Greenhouse Evaluations – Refined Rates

**Infestation:** 2 pairs of HBW

*Steinernema carpocapsae*

## **Rates:**

- 428,525 IJ/m<sup>2</sup> approx. (full rate)
- 75% of the full rate
- 50% of the full rate
- 25% of the full rate

sprayed at the foliage until runoff

N = 7 plants per treatment

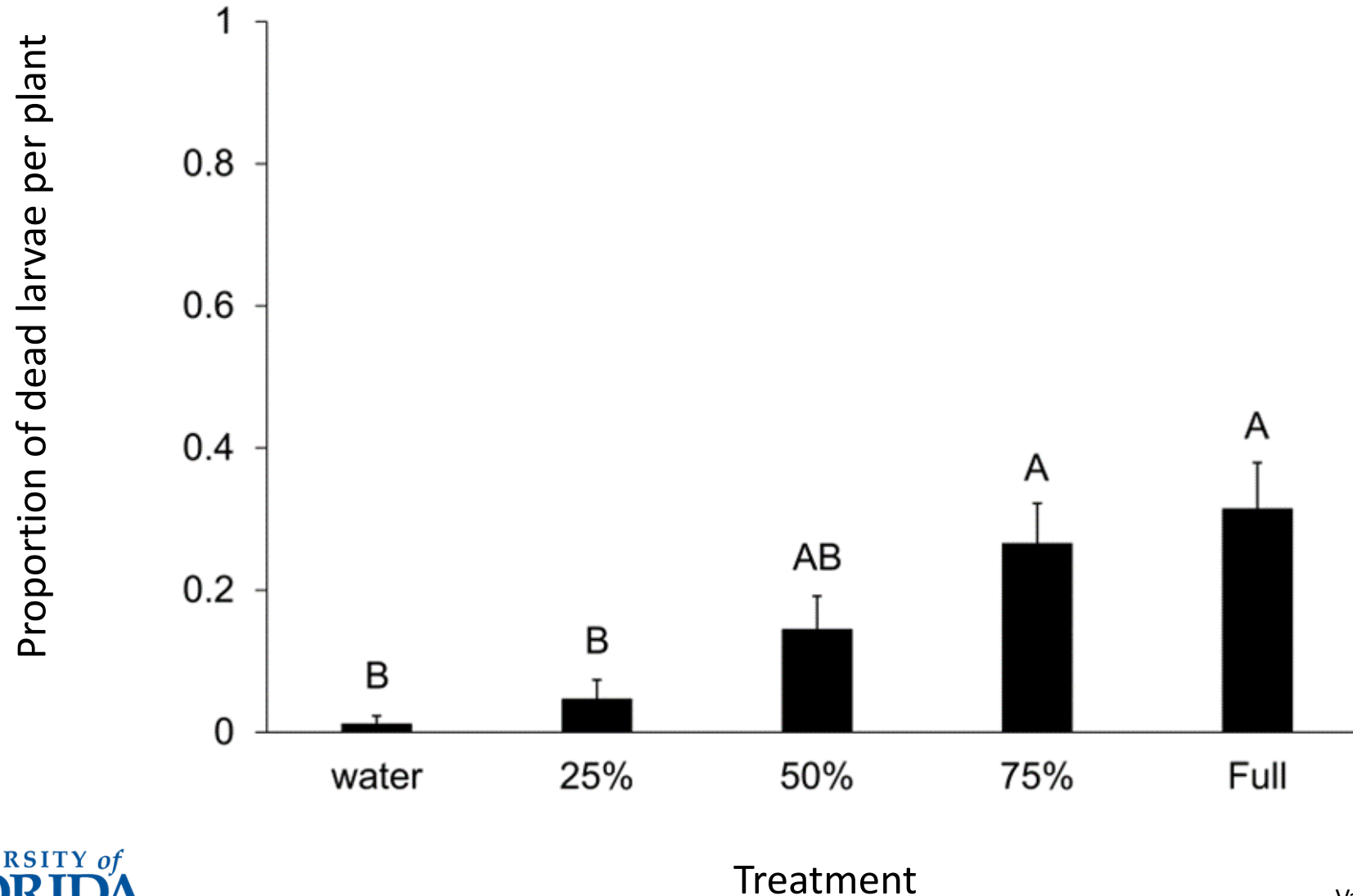
**Observations:** 5 actively growing buds and 5 fallen buds were dissected 1wk after the application



Photo: G. Vargas

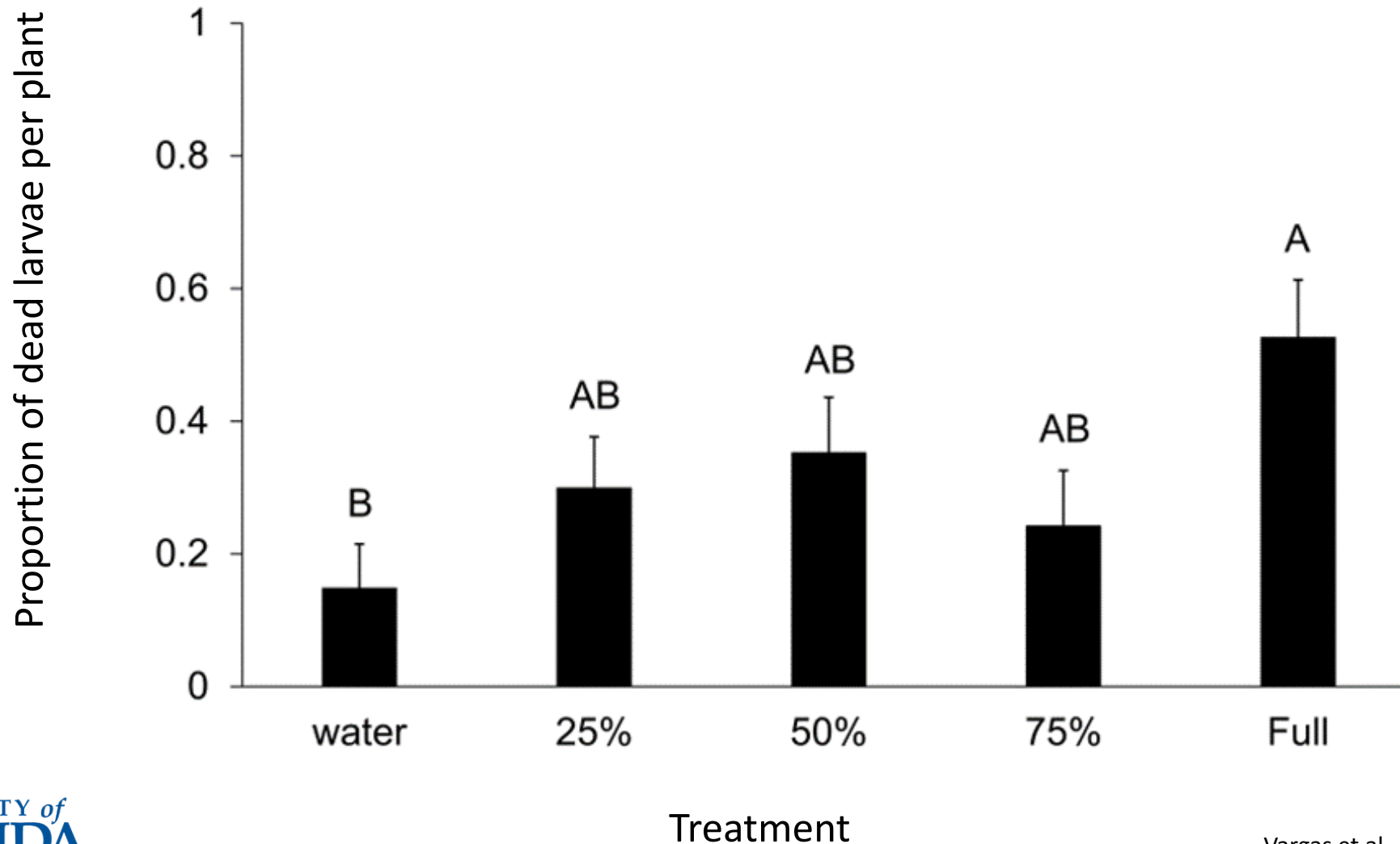
# Refined Rates of *S. carpocapsae* - Actively Growing Buds

Treatment: ( $\chi^2 = 23.09$ ;  $df = 4$ ;  $P < 0.001$ )



# Refined Rates of *S. carpocapsae* - Fallen Buds

Treatment: ( $\chi^2 = 11.33$ ;  $df = 4$ ;  $P = 0.023$ )



# EPNs- Conclusions

- *Steinernema carpocapsae* and *S. feltiae* → best HBW control
- *Steinernema carpocapsae* greater efficacy → canopy and soil surface
- A 25% reduction in *S. carpocapsae* rate did not significantly decrease the efficacy of controlling larvae at the canopy
- EPN application can target HBW larvae in fallen infested buds



Photo: G. Vargas

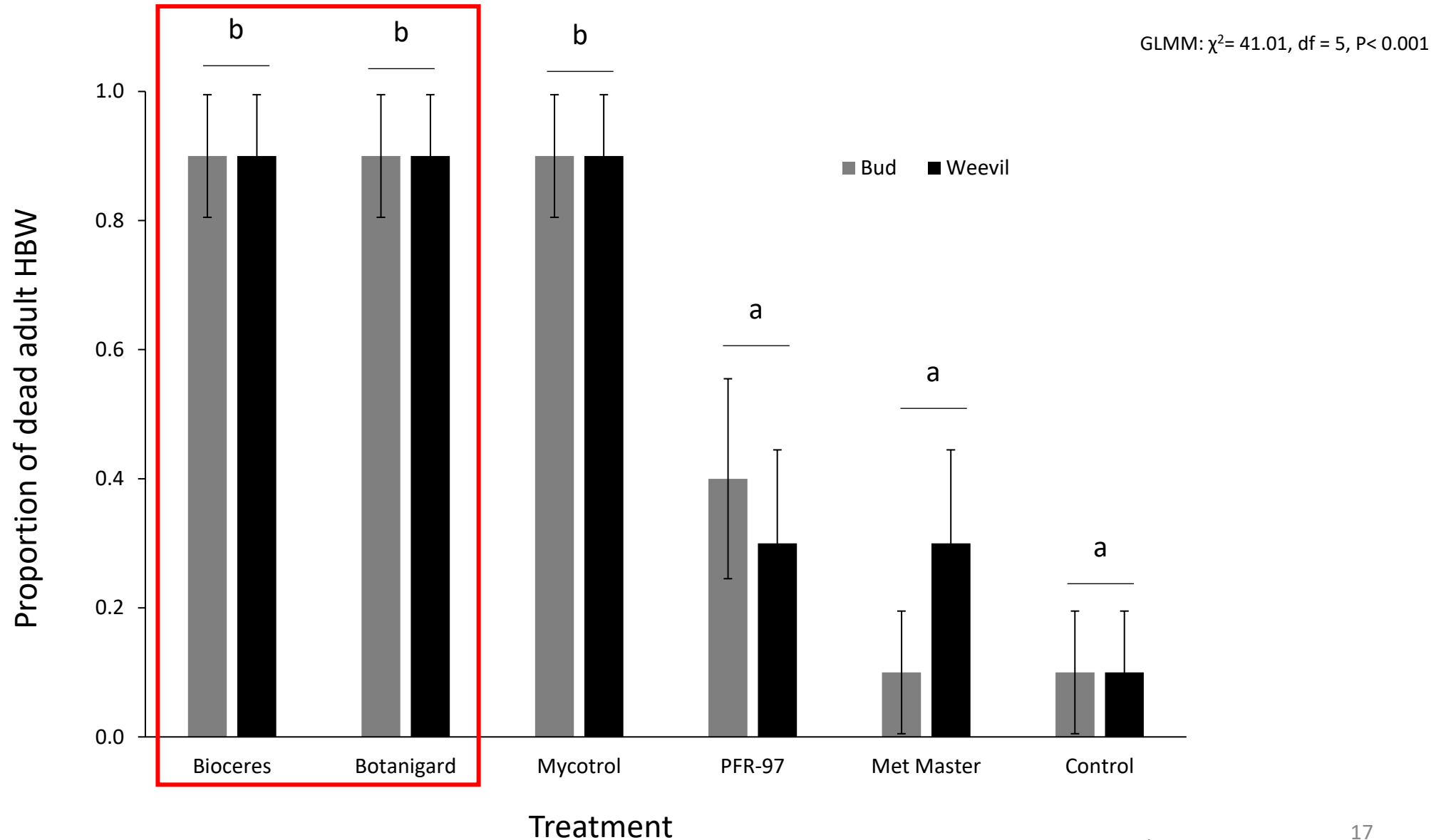
# Biological Control of HBW - Entomopathogenic Fungi

- Tested entomopathogenic fungi in lab assays against adult weevils
- Exposure through direct contact or treated buds
- Observation: 3-, 7-, 10-, and 14-days post treatment

Product	Active Ingredient	Rate
Bioceres WP	<i>Beauveria bassiana</i> , Strain ANT-03	4.8 x 10 <sup>7</sup> spores/ml
Botanigard 22WP	<i>Beauveria bassiana</i> , Strain GHA	3.4 x 10 <sup>7</sup> spores/ml
Mycotrol WPO	<i>Beauveria bassiana</i> , Strain GHA	4.1 x 10 <sup>7</sup> spores/ml
PFR-97 20% WDG	<i>Isaria fumosorosea</i> Apopka Strain 97	4.1 x 10 <sup>7</sup> spores/ml
Met52 EC	<i>Metarhizium anisopliae</i> Strain F52	3.7 x 10 <sup>7</sup> spores/ml
Control	Water	NA



# Laboratory Evaluations - Entomopathogenic Fungi





*Beauveria bassiana* ANT-03  
(BioCeres WP)



*Beauveria bassiana* Strain GHA  
(BotaniGard 22WP)



*Isaria fumosorosea*  
(PRF-97<sup>®</sup>WDG)



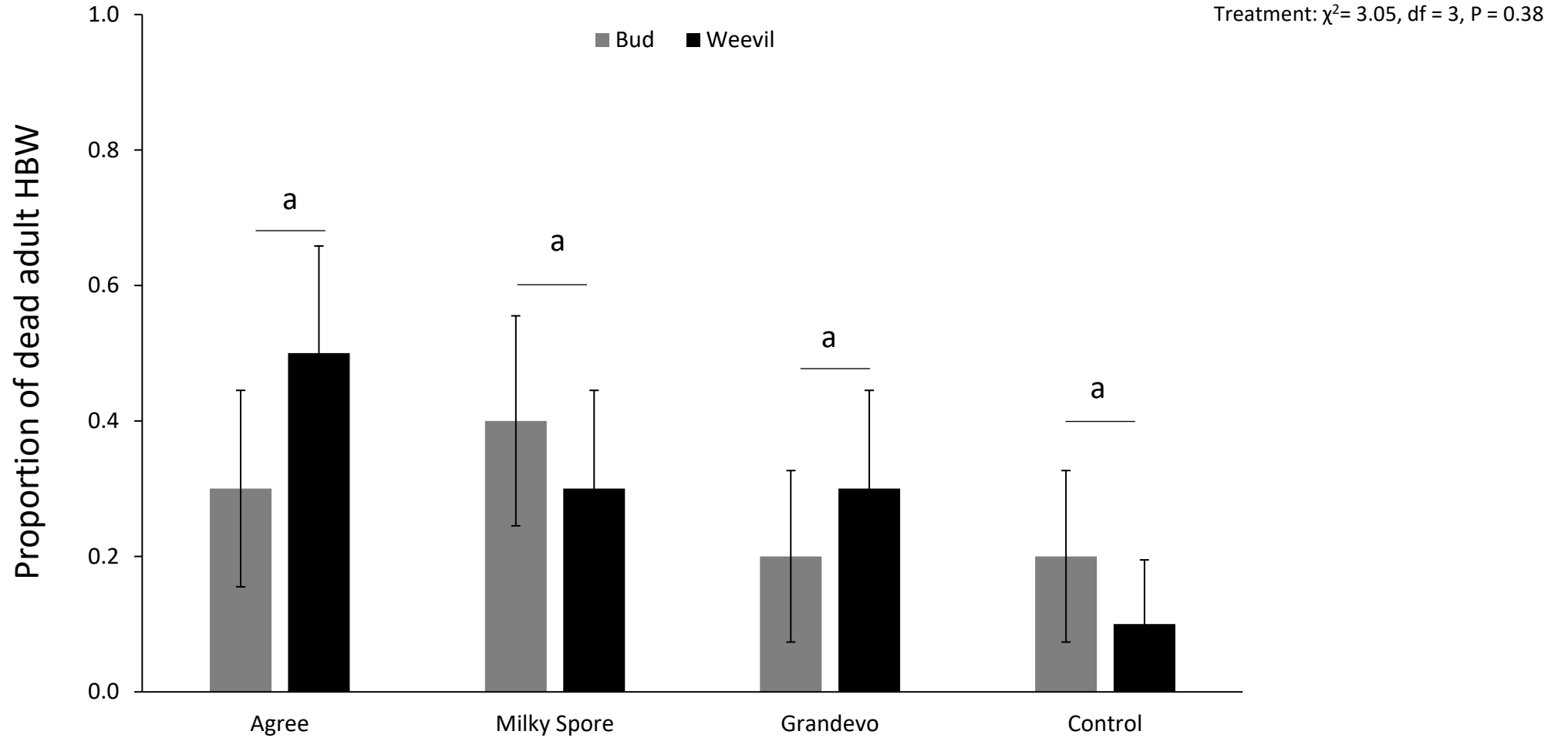
*Beauveria bassiana* Strain GHA  
(Mycotrol WPO)

# Biological Control of HBW - Entomopathogenic Bacteria

- Tested entomopathogenic bacteria in lab assays against adult weevils
- Exposure through direct contact or treated buds

Product	Active Ingredient	Rate
Agree WG	<i>Bacillus thuringiensis</i> subspecies <i>aizawai</i> strain GC-91	
Milky Spore	<i>Bacillus popilliae</i>	1 gr/200 ml
Grandevo	<i>Chromobacterium subtsugae</i> strain PRAA4-1 <sup>T</sup>	
Control	Water	NA

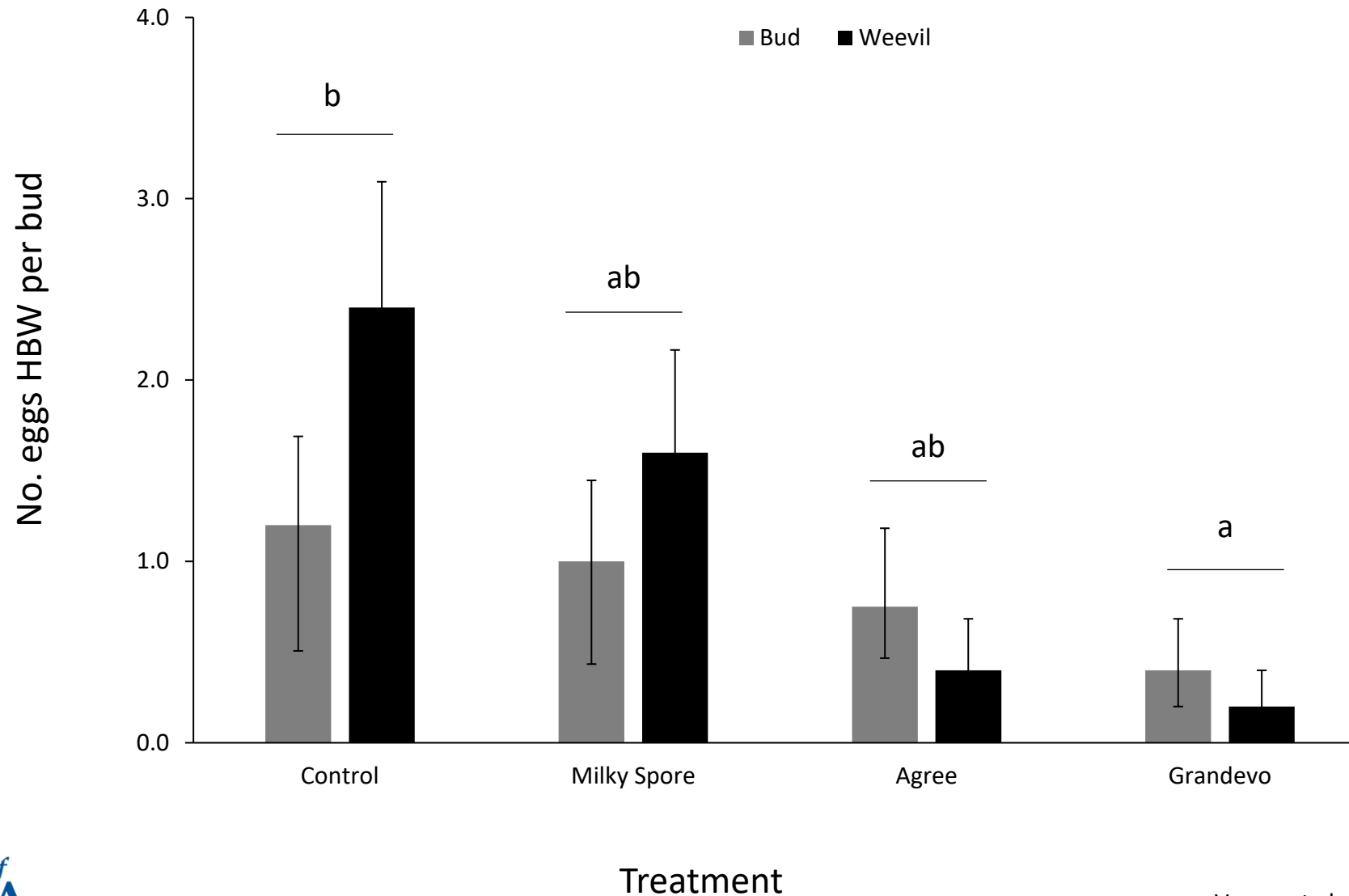
# Laboratory Evaluations - Entomopathogenic Bacteria



Treatment

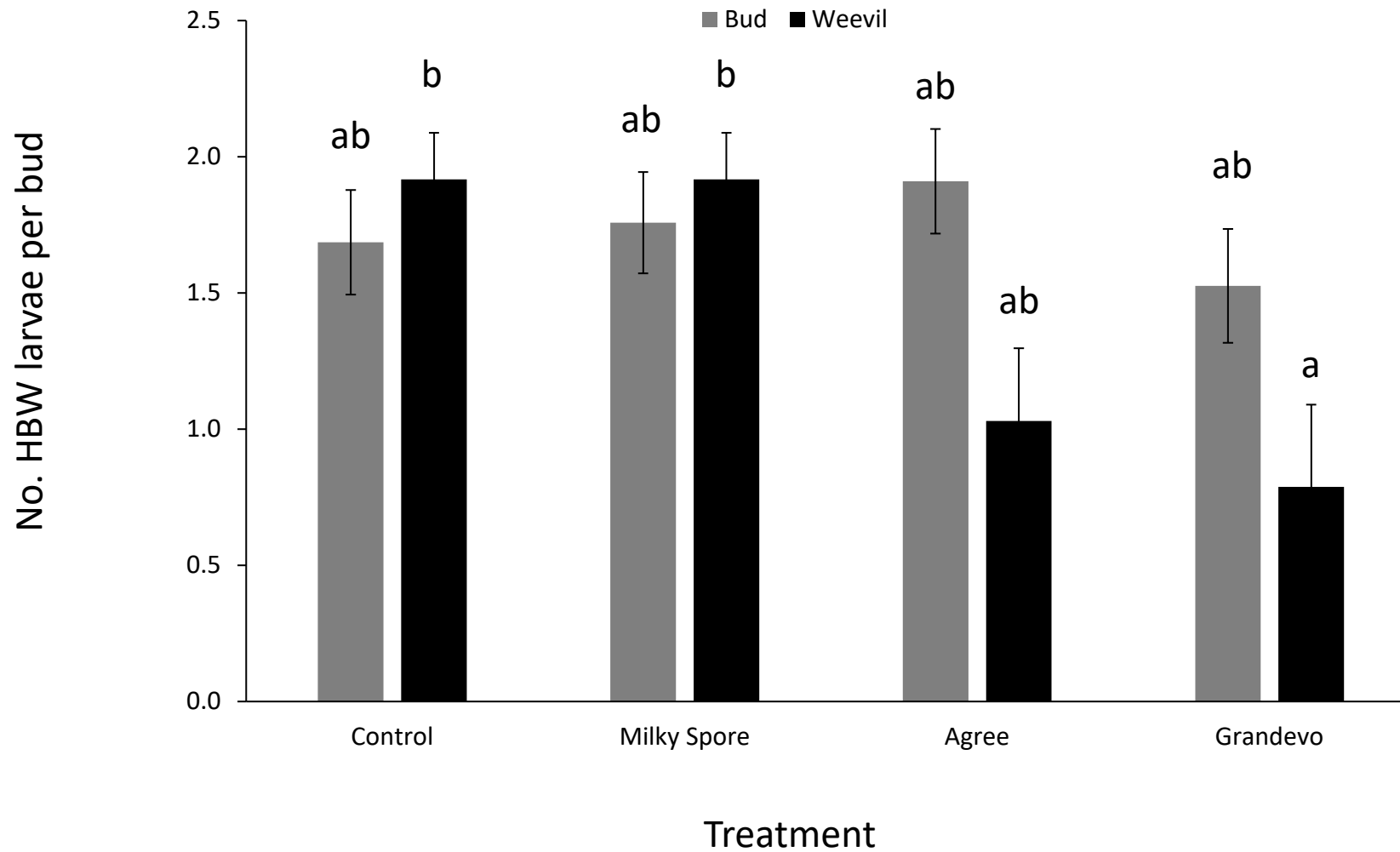
# Laboratory Evaluations - Entomopathogenic Bacteria

Treatment:  $\chi^2 = 9.71$ ,  $df = 3$ ,  $P = 0.02$



# Laboratory Evaluations - Entomopathogenic Bacteria

Treatment\*Contact:  
 $\chi^2 = 9.14$ ,  $df = 3$ ,  $P = 0.03$



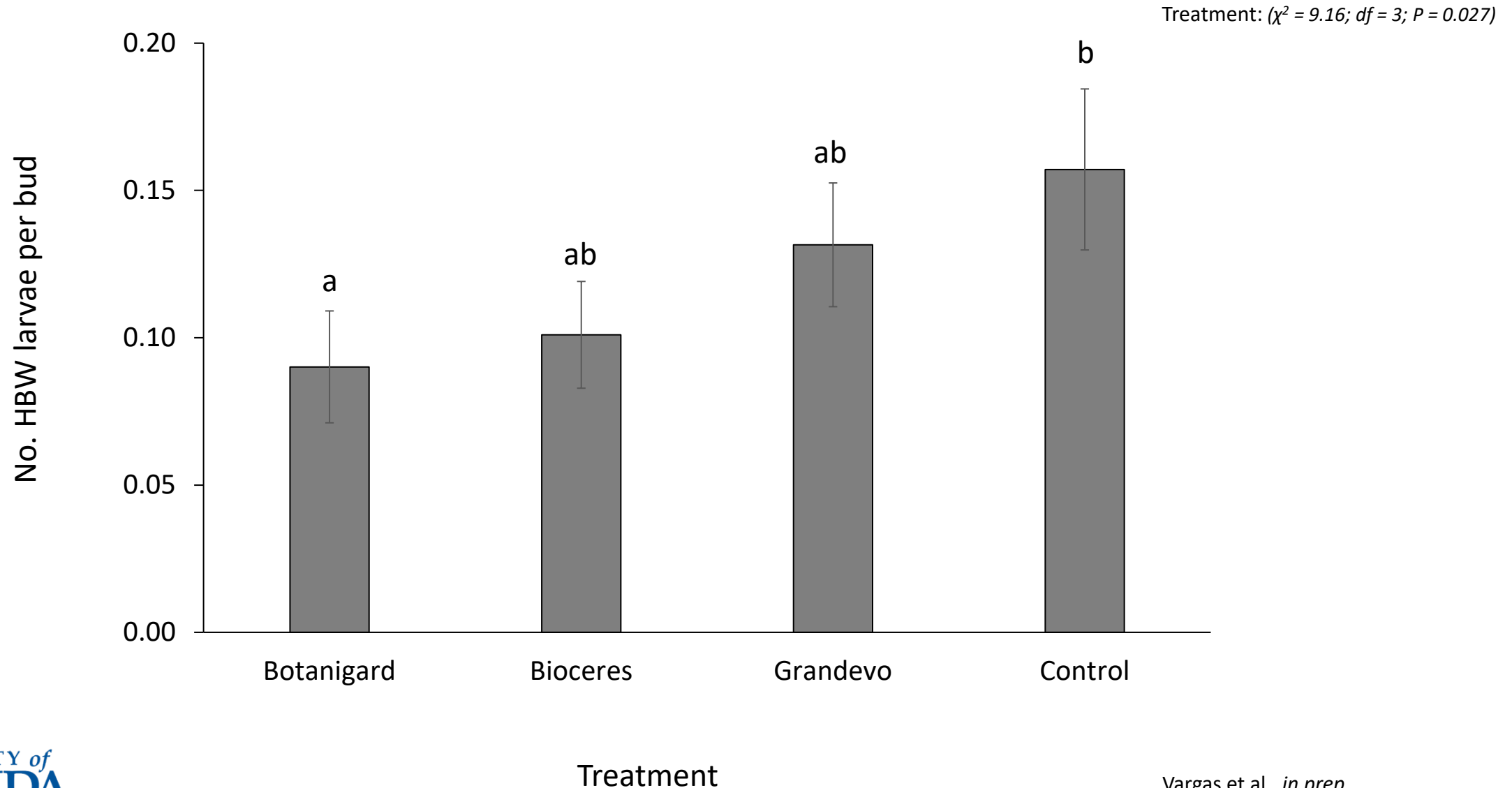
# Greenhouse Evaluations- Entomopathogenic Fungi & Bacteria

- Infestation: 2 HBW pairs/plant
- N = 9 plants/treatment
- Observation: 2 wks post application
- Collection of all buds (actively growing and fallen)
- 2 Blocks

Product	Active Ingredient	Rate
Bioceres WP	<i>Beauveria bassiana</i> , Strain ANT-03	3.59 gr/L
Botanigard 22 WP	<i>Beauveria bassiana</i> , Strain GHA	2.37 gr/L
Grandevo	<i>Chromobacterium subtsugae</i> strain PRAA4-1 <sup>T</sup>	20.7 gr/L
Control	Water	NA



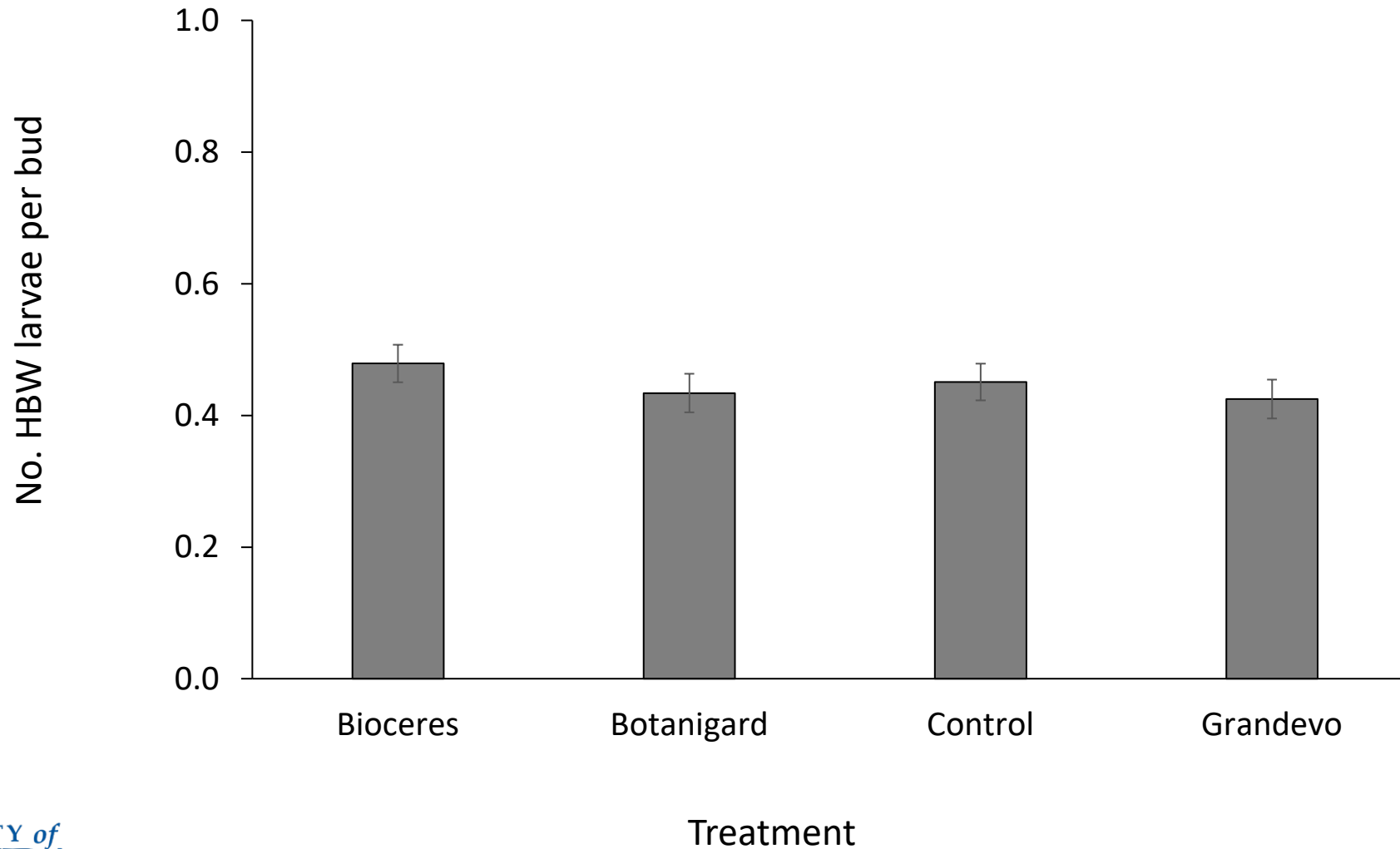
# Greenhouse Evaluations- Actively Growing Buds





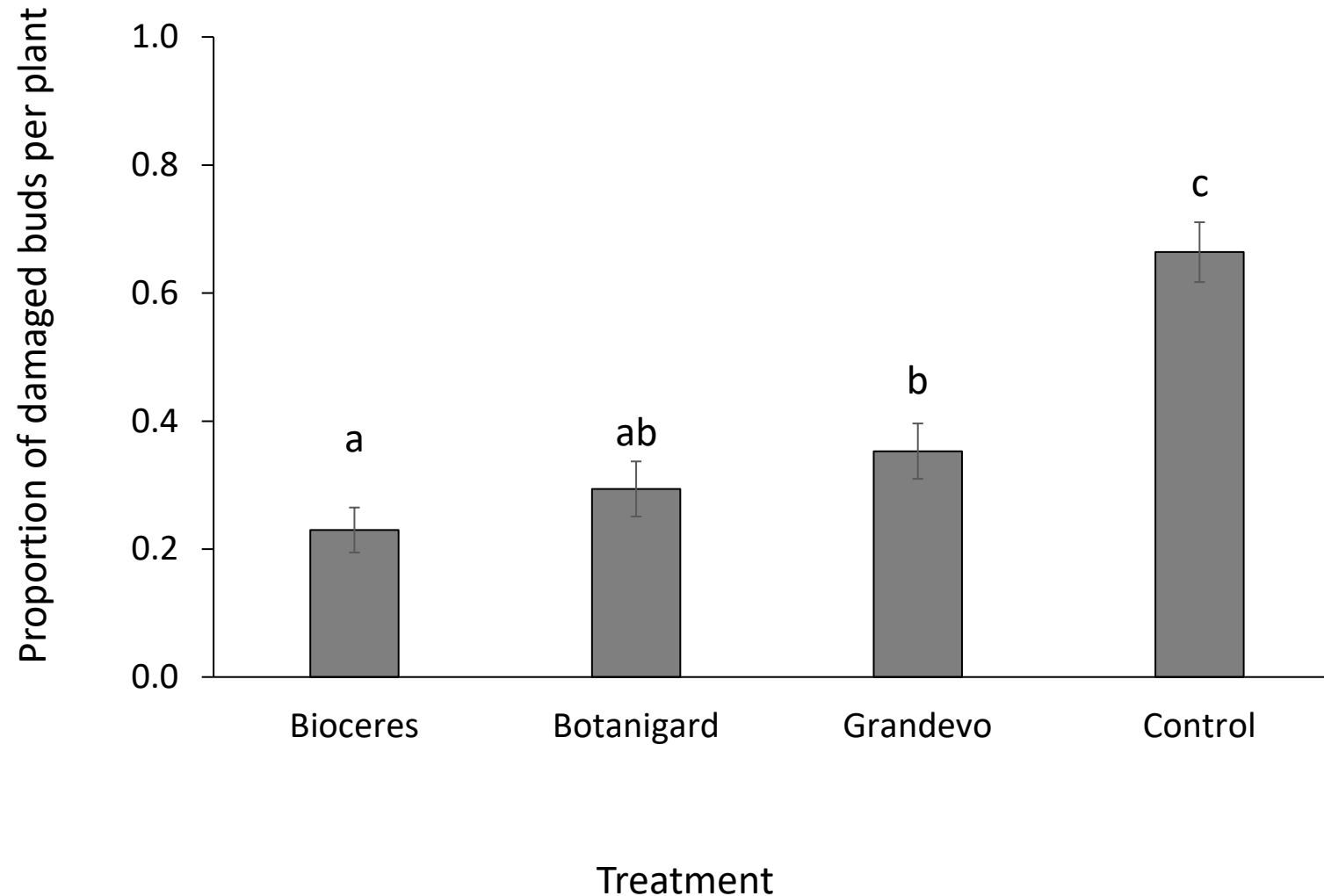
# Greenhouse Evaluations- Fallen Buds

Treatment: ( $\chi^2 = 5.38$ ;  $df = 3$ ;  $P = 0.15$ )



# Greenhouse Evaluations- Damage on Actively Growing Buds

Treatment: ( $\chi^2 = 121.07$ ;  $df = 3$ ;  $P < 0.001$ )



# Entomopathogenic Fungi & Bacteria- Conclusions

- *Both B. bassiana* strains → efficacious against the HBW
- Grandevo → possible antifeeding and anti-oviposition effects
- All products resulted in greater number of actively growing buds

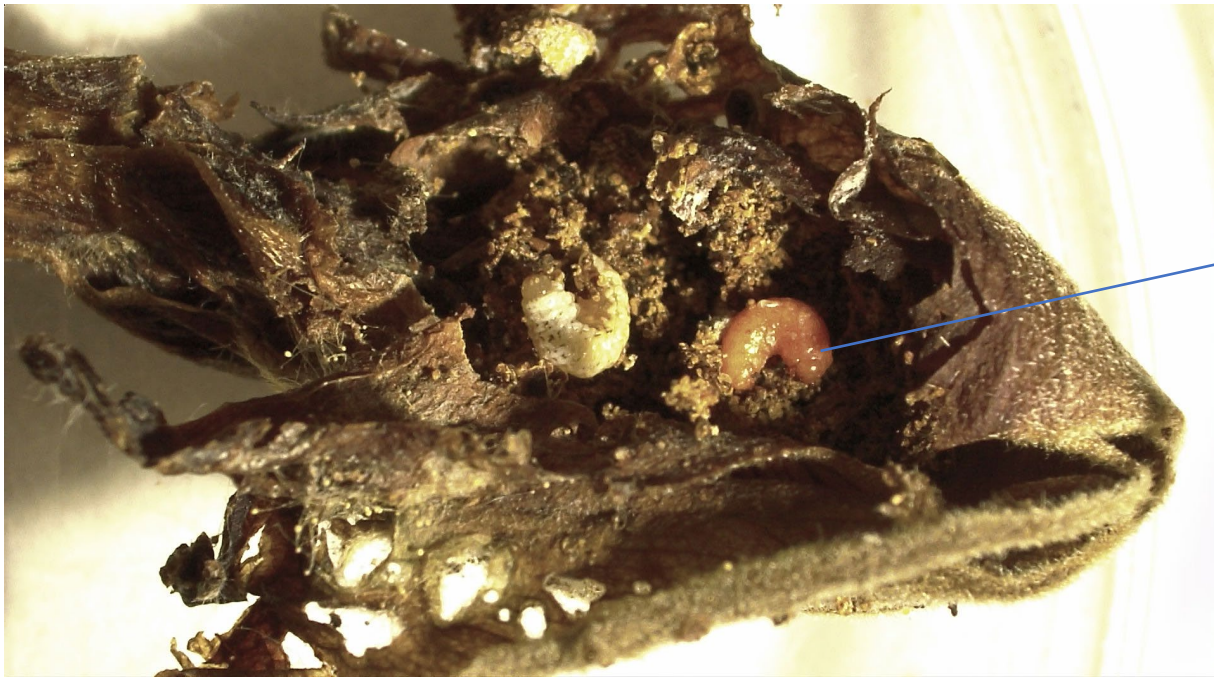


# Biological Control of HBW - Parasitoids

- *Catolaccus hunteri* (Hymenoptera: Pteromalidae)
- Polyphagous ectoparasitoid
- Parasitoid of several species of *Anthonomus* sp.
  - cotton boll weevil
  - pepper weevil
- It is now available in the US market



# *Catolaccus hunteri*



Parasitized larva



Photo: Y. Velazquez Hernandez

# *Catolaccus hunteri*

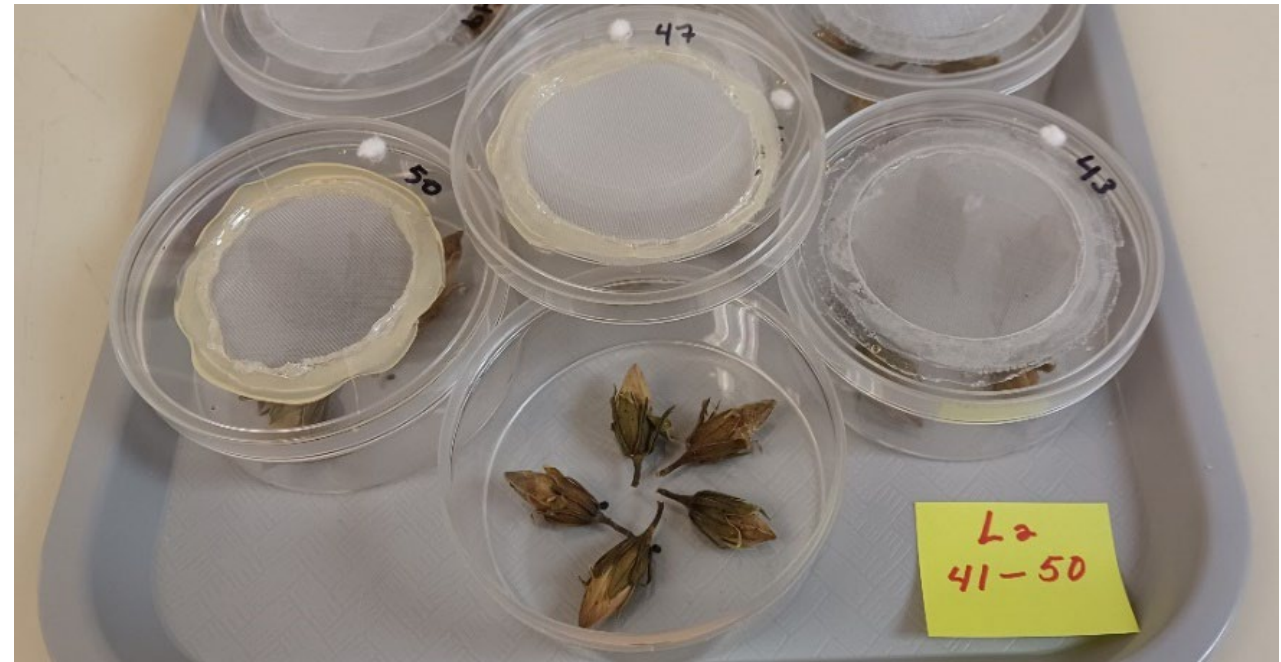
## Parasitized Pupa



Photo: Y. Velazquez Hernandez

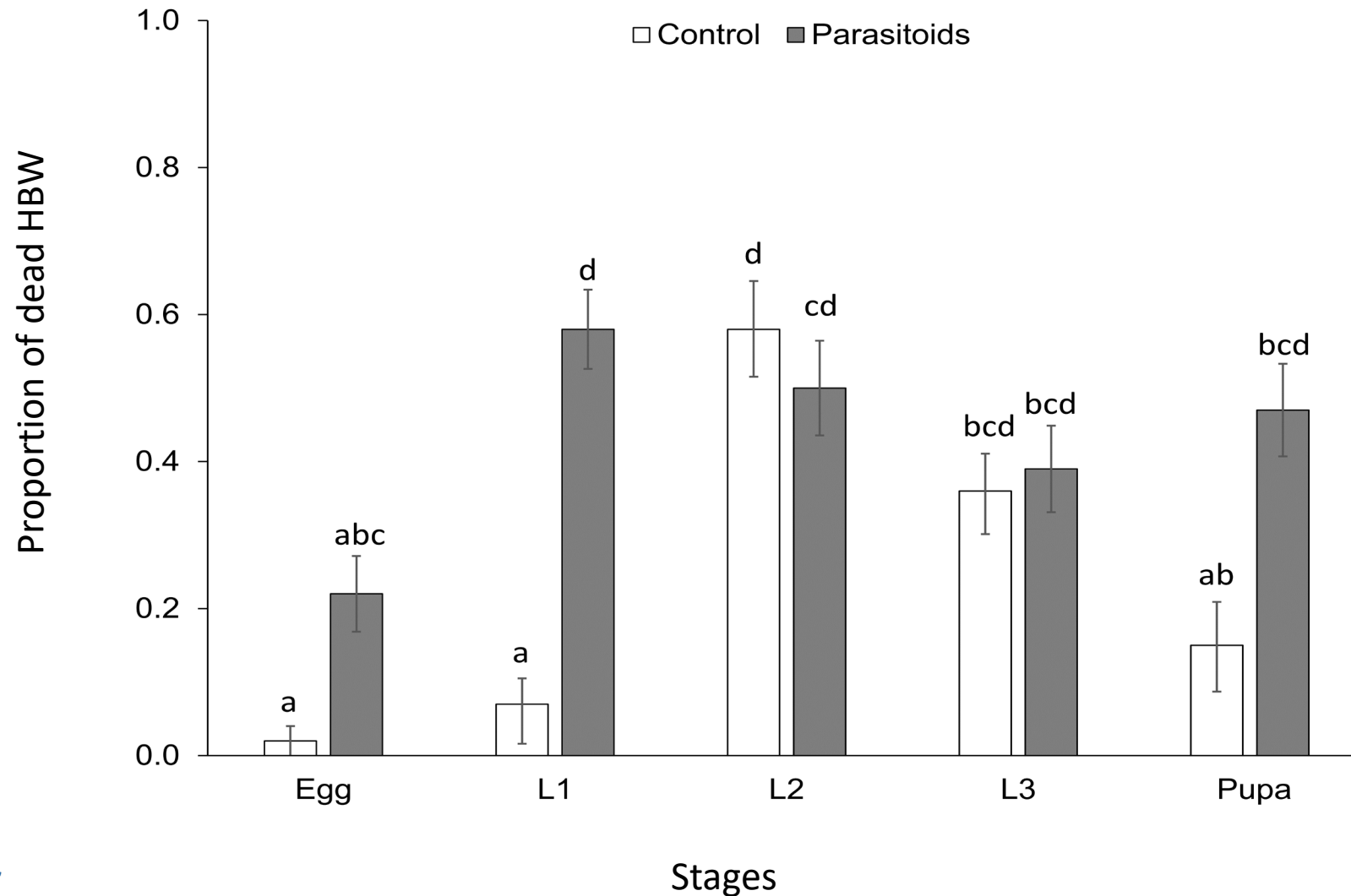
# Which is the most appropriate HBW developmental stage for *C. hunteri*?

- Lab evaluations
- 5 buds with:
  - Eggs
  - L1-L3
  - Pupa
- N = 10 per stage
- 1 parasitoid pair released for 24h



# Laboratory Evaluations- Parasitoids

Treatment\*Stage:  
 $\chi^2 = 32.99$ ; df = 4; P < 0.001





# Is there an Optimum Release Rate for the *C. hunteri*?

- Lab evaluations
- 15 HBW infested buds plus:
  - 1
  - 2
  - 3 female parasitoids
- N = 4 per treatment
- Parasitoids remained in the cage for 4 days

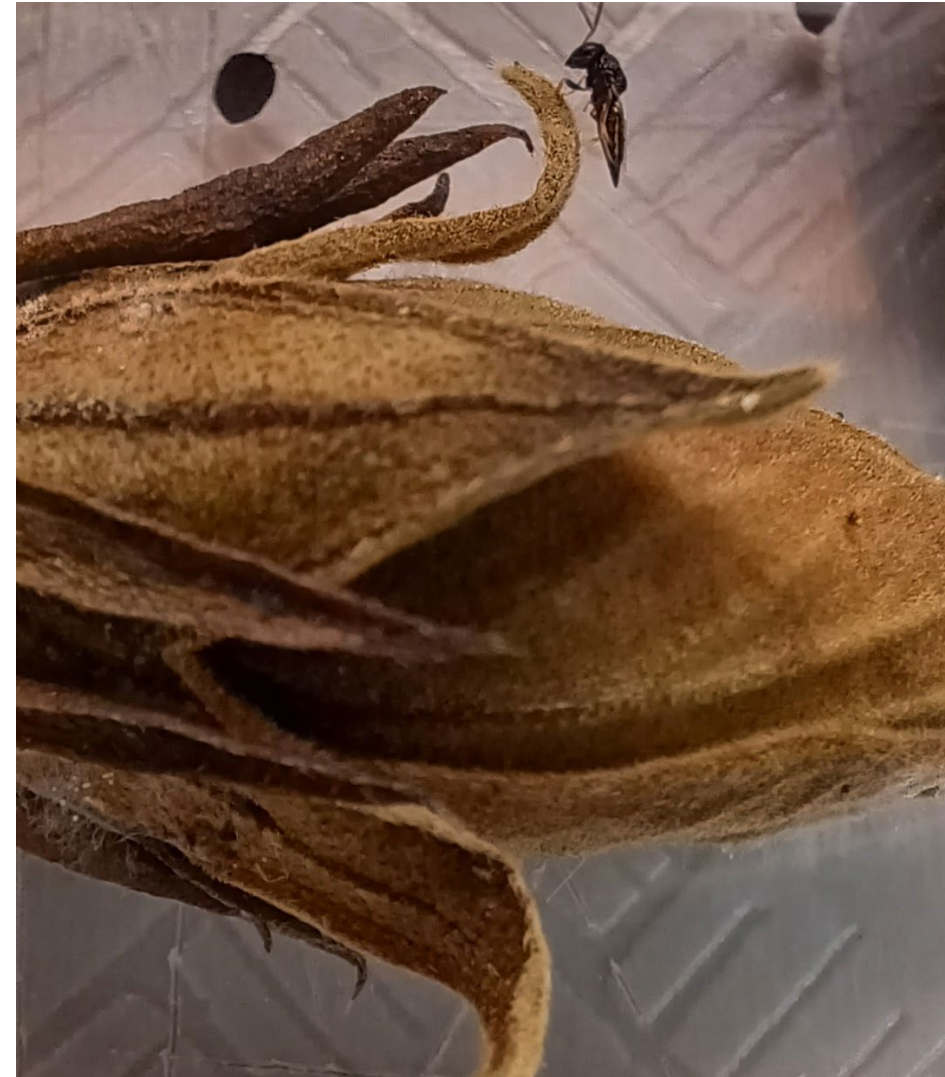
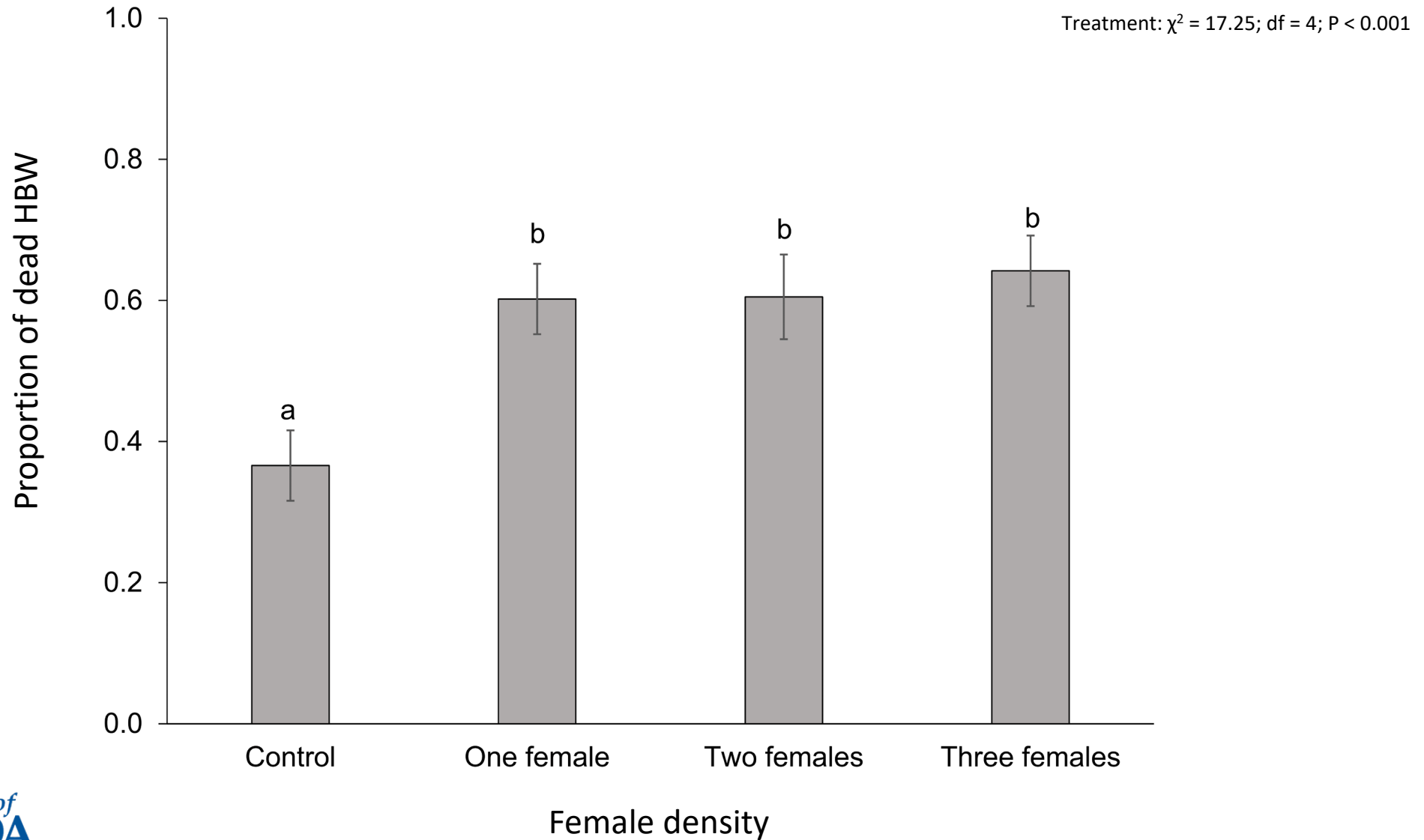


Photo: Y. Velazquez Hernandez

# Laboratory Evaluations- Parasitoid Rate



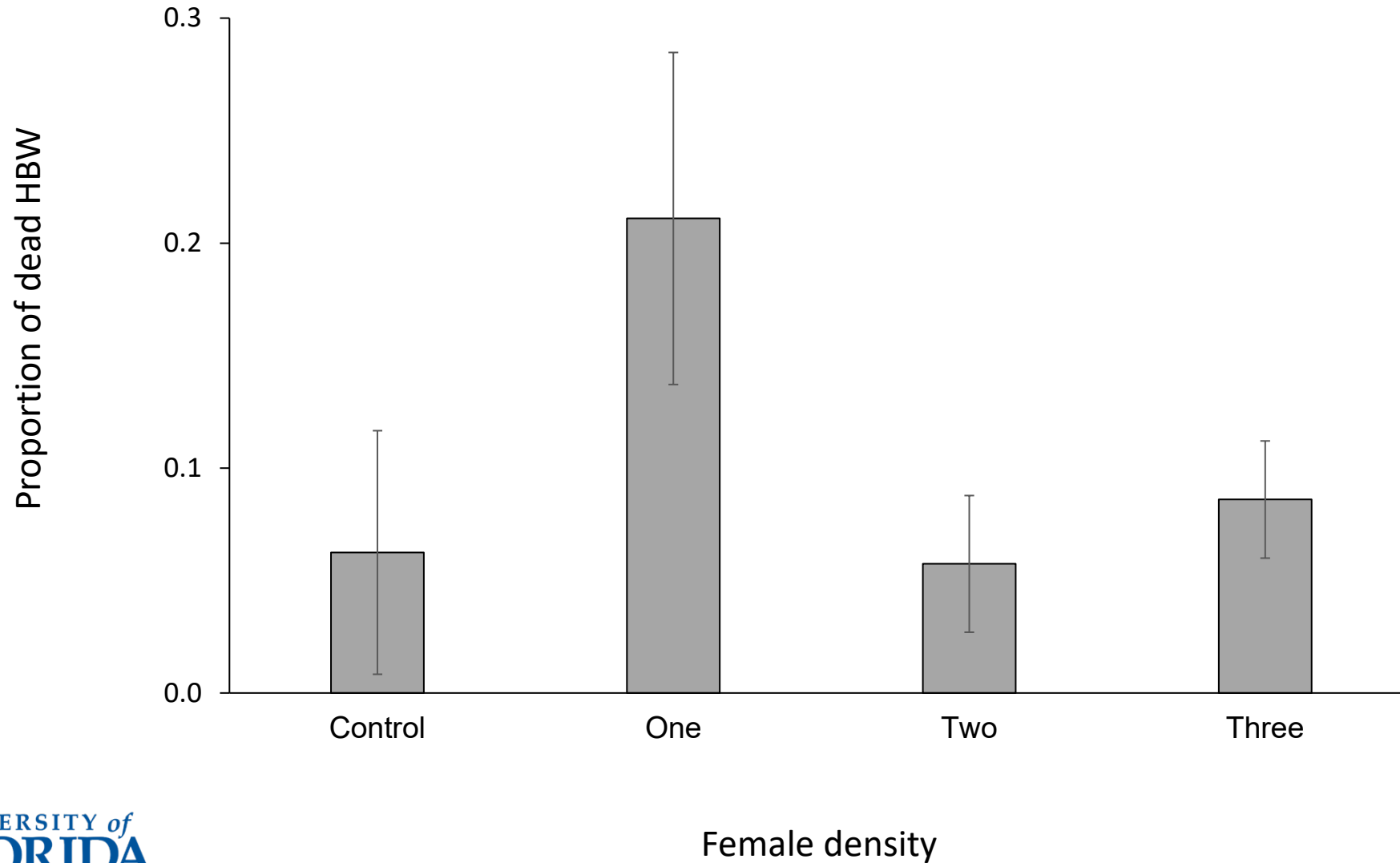
# Is there an Optimum Release Rate for the *C. hunteri*?

- Greenhouse evaluations
- Individual hibiscus infested plants plus:
  - 1
  - 2
  - 3 parasitoid pairs
- N = 4 per treatment
- Collection of all buds 2 wks post parasitoid release



Photo: Y. Velazquez Hernandez

# Greenhouse Evaluations- Parasitoid Rate



# Parasitoids- Conclusions

- *Catolaccus hunteri* shows potential in controlling HBW larvae
- Releasing 1 female per 15 flower buds, did not differ significantly from releasing 2-3 individuals, but caused more mortality than the control, under lab conditions
- A released of a single parasitoid pair showed potential to control HBW under greenhouse conditions
- More research to define optimum sex ratio and its effects on female's reproductive performance



Photo: Y. Velazquez Hernandez

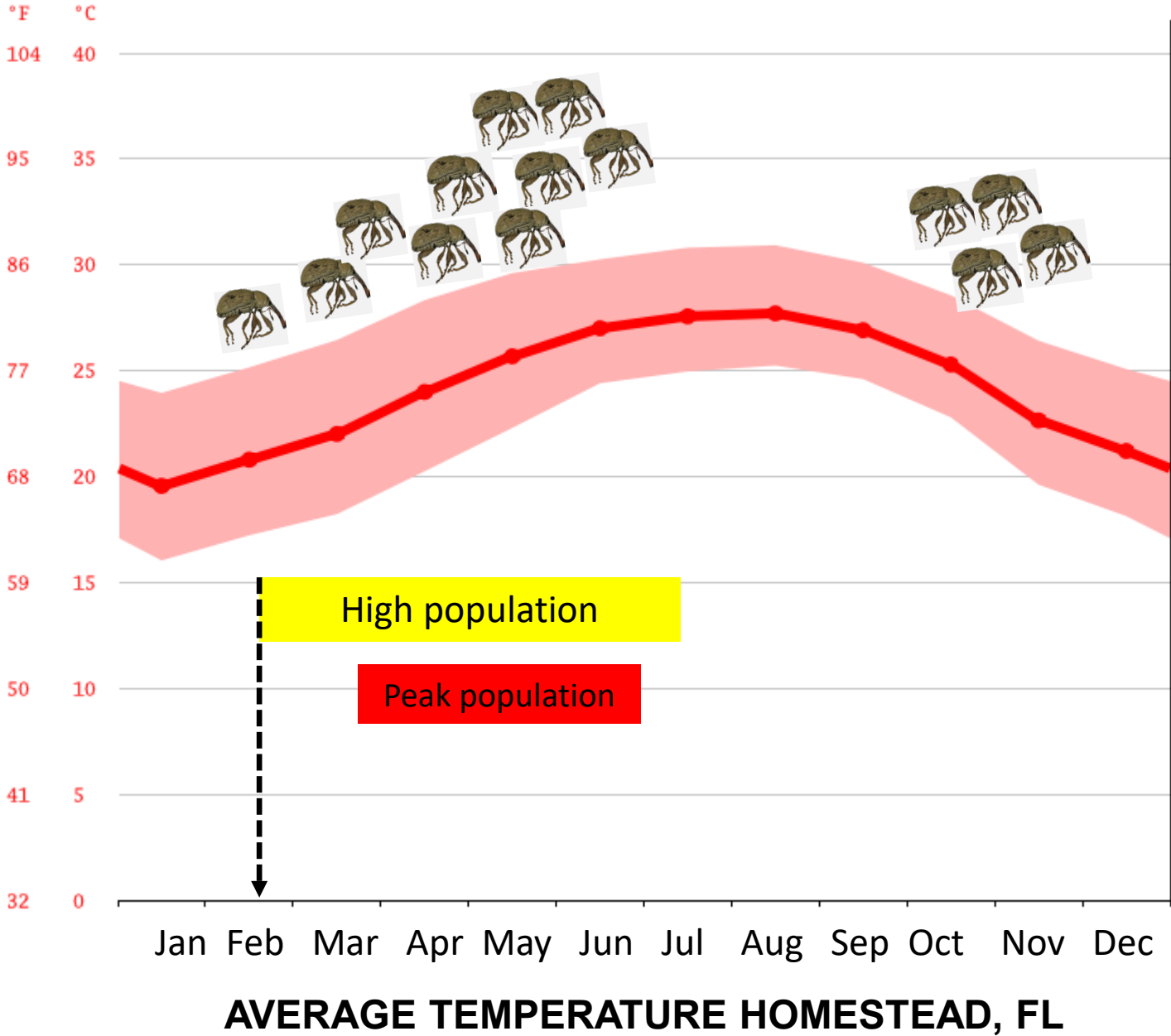
# Considering phenology towards an IPM program

Monitor and sanitize throughout the year

Systemics under a prophylactic approach from February

Use of foliar entomopathogenic nematodes, fungi and bacteria can help in sustaining low populations

Use of contact insecticides before the shipping period



<https://en.climate-data.org/north-america/usa/florida/homestead-1632/#temperature-graph>

# Biological Control of HBW – Take-home Messages

- *Steinernema carpocapsae* and *S. feltiae* → best HBW control
- *Beauveria bassiana* Strain ANT-03 and Strain GHA → efficacious against the HBW
- *Chromobacterium subtsugae* strain PRAA4-1<sup>T</sup> → viable alternative
- *Catolaccus hunteri* → potential in controlling HBW larvae, more research is needed

# Sanitation

- The collection and destruction of fallen buds
- Reduces HBW population
- Environmentally friendly strategy
- Labor intensive



Photo: A.D. Greene



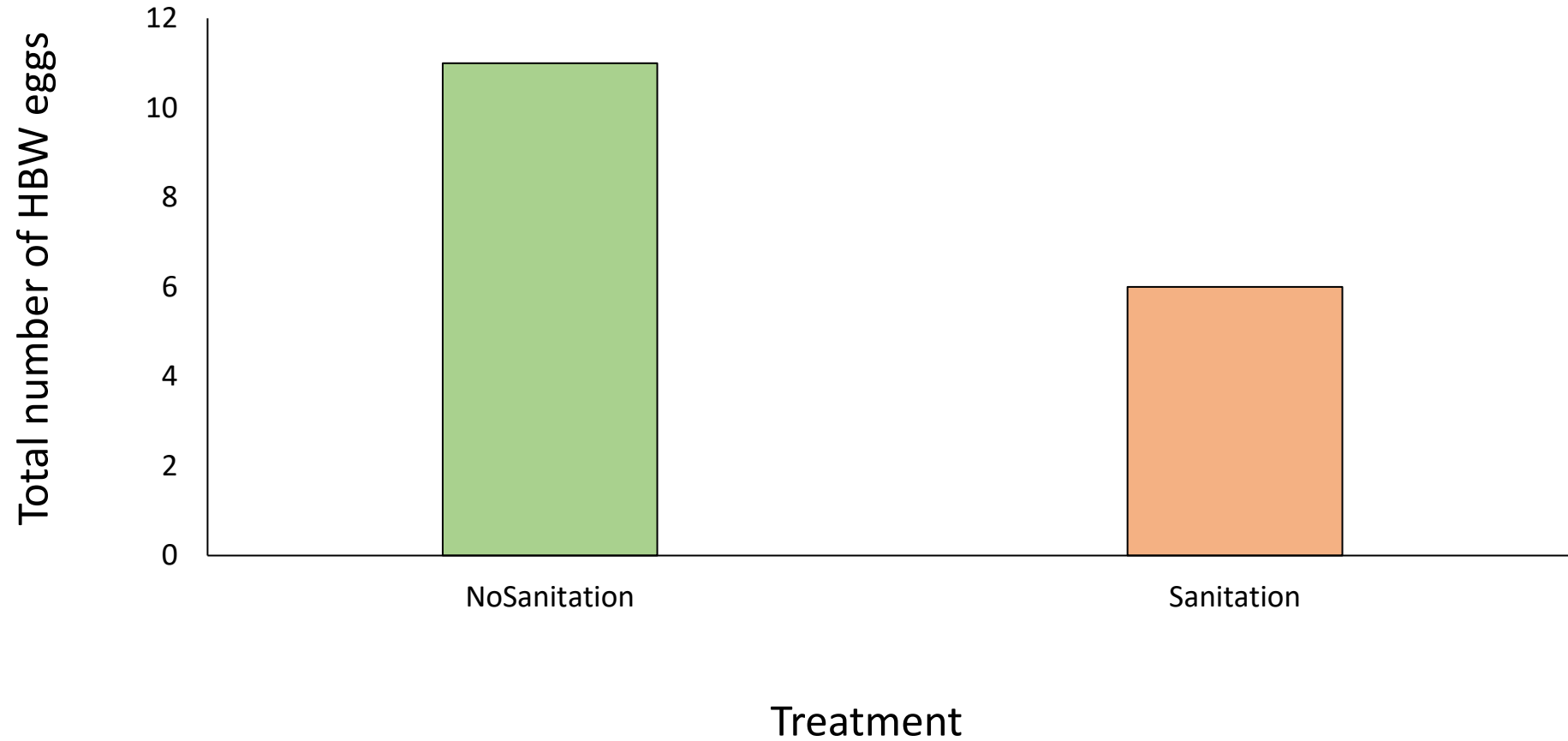
# Effect of Sanitation on HBW Populations

- 4 tents → Sanitation
- 4 tents → No Sanitation
- 10 hibiscus/tent
- Released 20 HBW (10F/10M)/tent
- Sampled 5 actively growing buds weekly for 9 weeks plus fallen buds (sanitation)

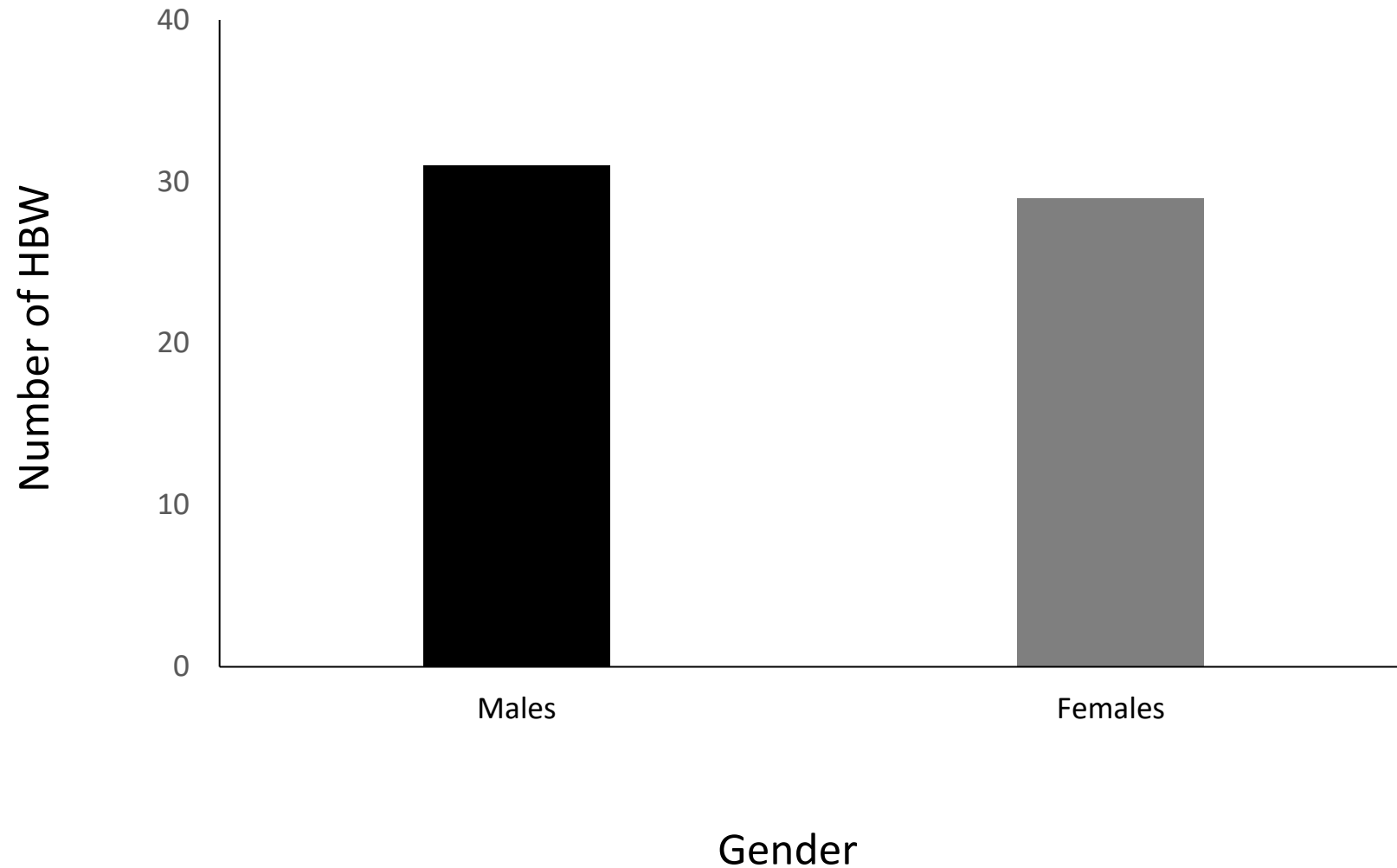


Photo: A.D. Greene

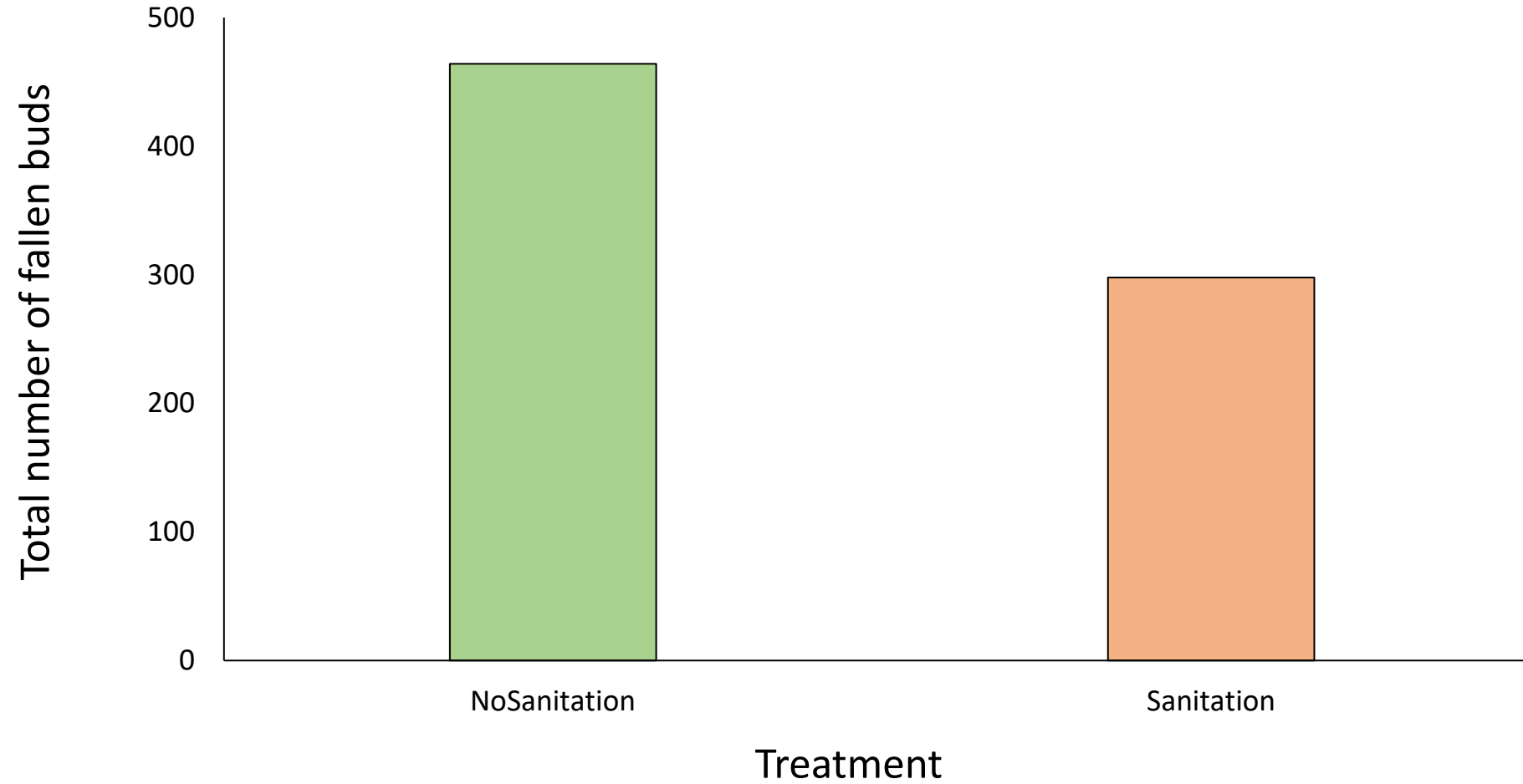
# Sanitation - HBW Eggs from Actively Growing Buds



# Sanitation - Emerging HBW from Fallen Buds



# Sanitation - Number of Fallen Buds



# Can We Optimize Sanitation?

- Increase:
  1. Speed
  2. Number of collected buds
- Options:
  1. Manual sanitation
  2. Leaf blower
  3. Vacuum

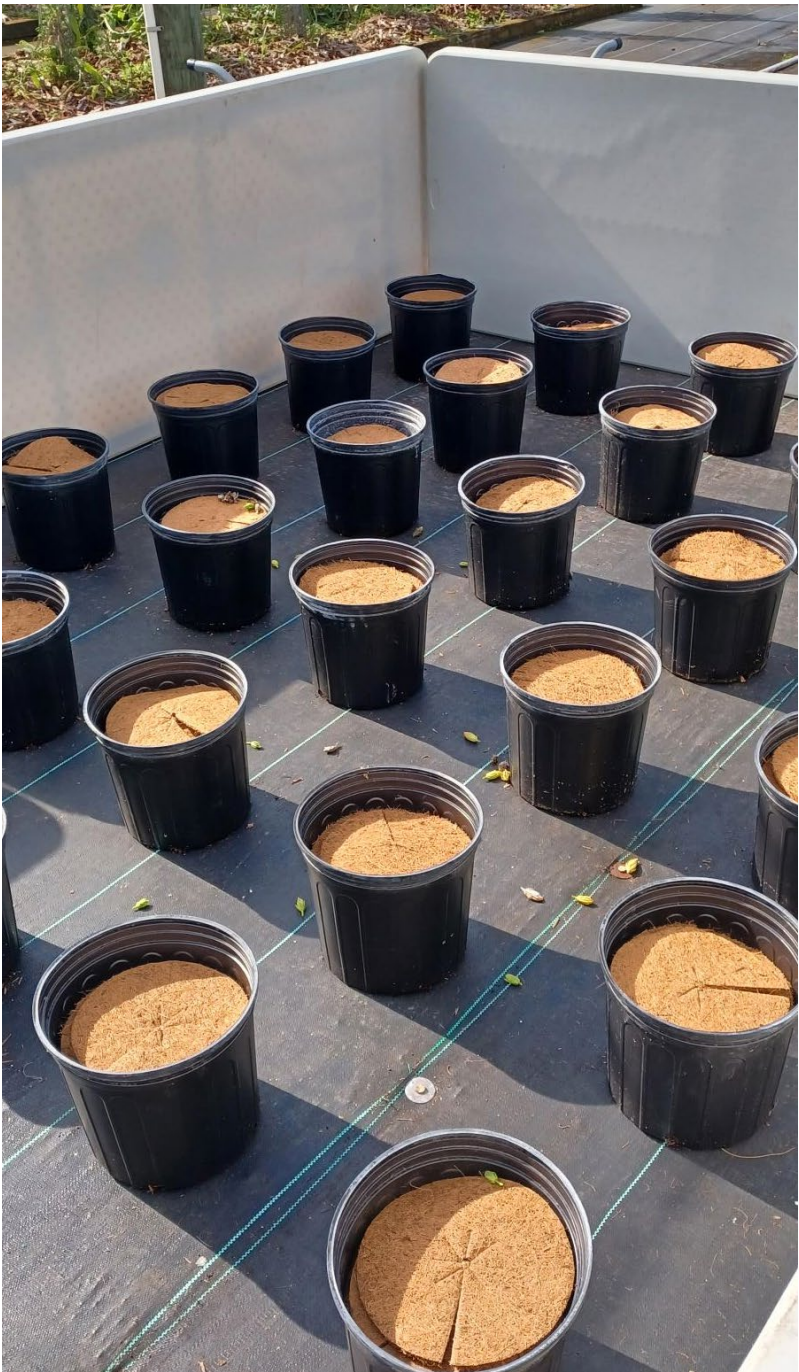


# Can We Optimize Sanitation?

- Experimental area: 12 ft x 6 ft
- 25 different aged infested buds (fresh, 1 and 2 wks)
- 50 pots (3 gal) +75 buds
- N = 10 per collection method

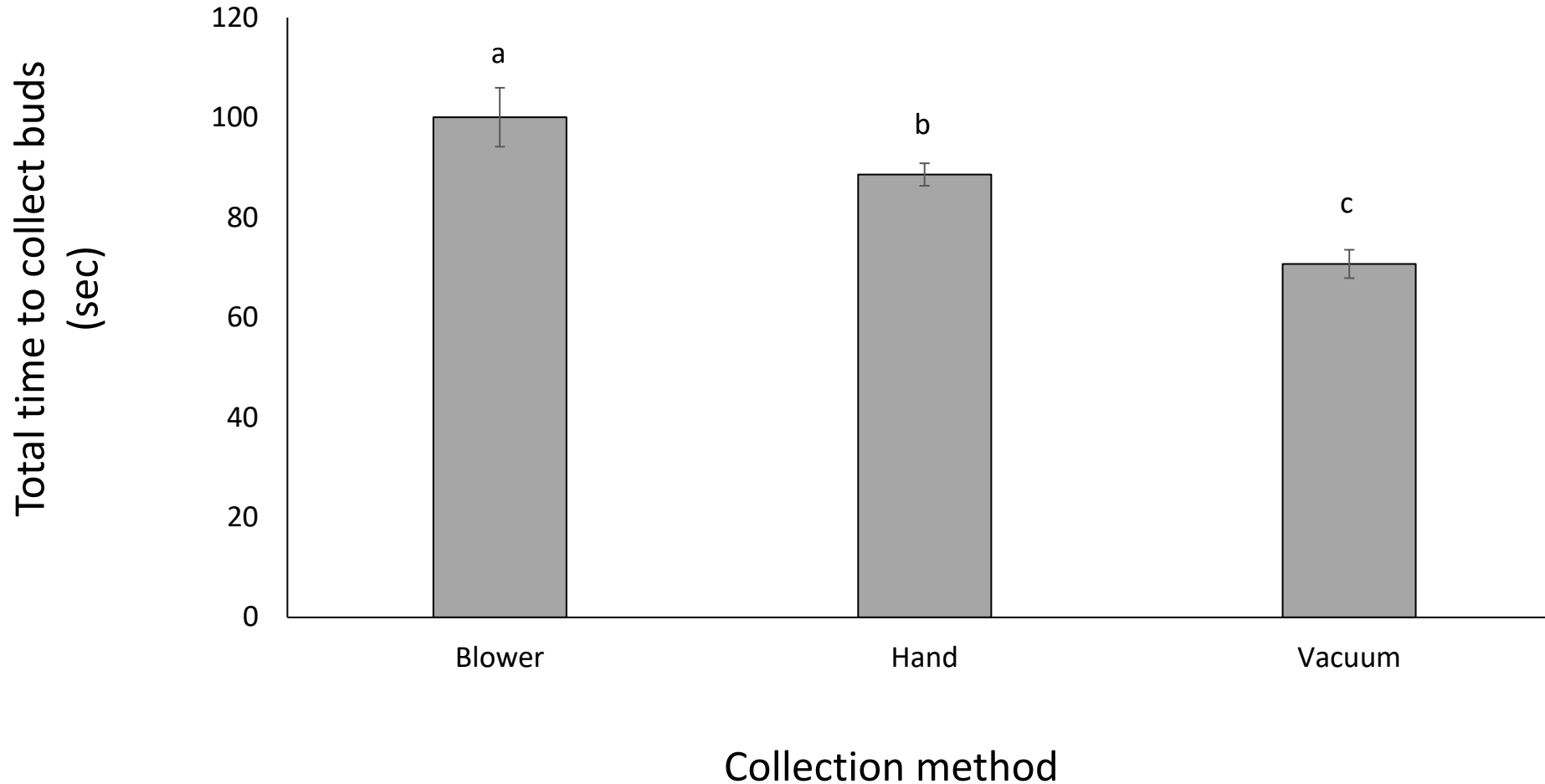


Photo: A.D. Greene



Photos: A.D. Greene

# Can We Optimize Sanitation?





# Sanitation - Conclusions

- Sanitation experiments revealed:
  - a reduction in fallen buds
  - a reduction in HBW eggs and larvae on healthy buds
  - prevention of at least 1.5 HBW/plant from reinfesting hibiscus plants
- Vacuum is the fastest method to implement sanitation



Photo: Y. Velazquez Hernandez

# Next Steps

- Continue research on biological control (parasitoids)
- Focus on chemical and behavioral ecology → develop a HBW lure
- Define variety preference



500µm

# Thank you!

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